# TFF-Easy: Tangential flow filtration for Extracellular Vesicle concentration



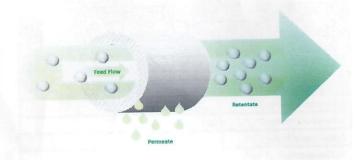


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About TFF-Easy.

TFF-Easy is a filter cartridge containing polysulfone hollow fibers (20 nm pores), which allows the concentration and the removal of small proteins and molecules from diluted matrices (cell conditioned media, urine, etc..), prior to the EV purification.

Water and small molecules (< 20 nm) pass through the hollow fiber pores, whereas EVs are concentrated in the retentate. EVs can be easily recovered with a syringe from the filter cartridge.



#### Specification.



1	Sample injection nozzle	
)	Tangential flow filtration nozzle	
3	Flow valve 1	
4	Permeate nozzles	
5	Tangential flow filtration nozzle	
6	Flow valve 2	

Sample	Volume range	Reusable	
Cell media	5 - 1000 ml	20-30 times	
Urine *	5- 1000 ml	10-20 times	
Plasma/serum	5-20 ml	5-10 times	

Fig 1

\* TFF-Easy can be used for concentrating all diluted body fluids (urine, CSF, bronchoalveolar lavage etc).

Sterility: The TFF-Easy is provided sterile. Once used the filter can be sterilised by Beta irradiation. Do not autoclave the TFF-Easy.

#### Storage.

Store the device at room temperature.

## EV concentration procedure.

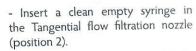
## 1- Sample injection by filtration (optional).



- Remove the screw from the sample injection nozzle (position 1).
- Insert a syringe filter in the nozzle. The device is compatible with multiple filter typology. The dimension of the filter pores can be choosen by the user according to the necessity.

In order to keep all the EV subtypes it is recommended to use filter pores larger than 0.22 µm.

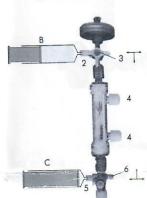
- Pick up the sample with syringe A and insert on the top of the filter.



- Rotate the valve 3 to the position indicated by arrows in figure 3.
- Inject the sample into the filter by pushing the piston of syringe A. The sample passes through the filter, filling the syringe B.

Fig 3

### 2- Tangential Flow Filtration.



- Rotate the valve 3 to the position indicated in figure 4.
- Open the permeate nozzles, removing the screws.
- Insert a new clean syringe (C) in position 5.
- Set 2 permeate collection tubes under the nozzles in position 4, as indicated in figure 4.

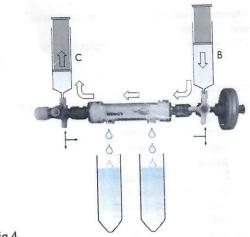


Fig 4

- Start the concentration process by pushing the syringes B and C alternatively upwards and downv.
- Continue the concentration procountil the desired volume is obtained.
- The permeate starts to flow to the . llection tubes, while the retentate in the syringes contains the conce. 'rated EVs.

## 3- Recovery of concentrated EVs.

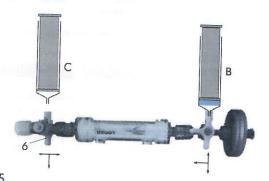


Fig 5

- In order to collect the concentrated EVs, push the concentrated products into syringe B.
- Rotate the valve 6 in the position indicate in figure 5 and disconnect the syringe C.
- Load air into syringe C.

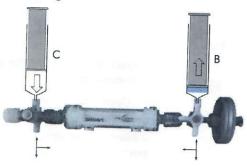


Fig 6

- Rotate the valve 6 to the position indicated in figure 6.
- Insert the syringe C and inject air into the device and pull up the piston of syringe B in order to collect all the residual volume of concentrated sample.



- Rotate the valve 3 to the position indicated by arrows and disconnect the syringe B.
- Collect the concentrated samples in a clean tube.

## Washing procedure.

Once the concentration process is ended the filter cartridge has to be washed with MilliQ water \*.

- Use 2 new clean syringes and load one of those with MilliQ water.
- Connect the syringe containing water to the position 2 and the empty one to the position 5
- Rotate the valve 3 and 6 in order to open the device (see fig 6)
- Inject the water in the device and continue the washing step by pushing alternatively the two syringes upwards and downwards until all the water is passed to the permeate collection tubes.
- Repeat the operation two times more.
- Let the device dry at room temperature.
- \* If used for concentrating complex fluids, the device can be washed with NaOH solution, followed by washing with MilliQ water.

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## Fast dialysis procedure.

TFF-Easy can be used for EV dialysis and buffer exchange.

- Inject the buffer (buffer 1) containing EVs into the device as indicated in figure 4.
- Push the syringes B and C alternatively upwards and downwards until all buffer has passed to the permeate collection tubes.
- Load the syringe B with the new buffer (buffer 2) and inject into the device. Repeat the operation indicated in the point above at least 5 times.
- Concentrate the EVs in buffer 2 until the desired volume.

#### Example

Dialysis progress	Conductivity (µS/cm)	Particle concentration (particle number/ml)
EVs in buffer 1 (PBS 1X) 5 ml	15000	5.8x10 <sup>11</sup>
1-Removal of buffer 1 by TFF	15000	
2- Injection of buffer 2 (NaCl 100 mM) in TFF cartridge		
3- Removal of buffer 2 and buffer 1 residues	4100	
4- Injection of buffer 2		
5- Removal of buffer 2	624	
6- Injection of buffer 2		
7- Concentration of buffer 2 up to 5 ml	621	4.9x10 <sup>11</sup>



